

RNA Isolation for qPCR

- Collect embryos or tissues in 1.5ml Eppendorf tube & remove excess media
- Add 10 volumes homogenization buffer (~250µl), triturate tissue to dissociate.
 - Homogenization Buffer:
 - 50mM NaCl
 - 50mM Tris-Cl (pH 7.5)
 - 5mM EDTA (pH 8.0)
 - 0.5% SDS
 - 200µg/ml proteinase K (thaw on ice until ready to add to HB)
- Incubate homogenate for 1hr at 37°C
- Add an equal volume of phenol:chloroform, vortex vigorously for 1'
- Spin at 5,000g for 10' at RT
- Transfer the aqueous phase to a clean tube and repeat phenol:chloroform extraction
- Transfer aqueous phase to clean tube and add 0.1 volumes of 3M sodium acetate (pH 5.2) and mix well.
- Add 2.5 volumes ice cold 100% ethanol, then incubate on ice for 2hrs
- Centrifuge at 5000g for 15' at 4°C.
- Remove supernatant and allow pellet to dry at RT
- Resuspend pellet in 20-50µl DEPC H₂O, then add an equal volume of 8M LiCl.
- Incubate at -20° C for at least 3hrs to o/n.
- Centrifuge at 10,000g for 30' 4°C.
- Remove supe, then wash pellet with 70% ethanol, centrifuge 5' as above.
- Remove supe and air dry pellet.
- Resuspend nucleic acids in small volume of DEPC H₂O.
- Run 1µl of nucleic acid on 1% check gel.
- DNase treat sample in 100µl volume
 - Example: 14µl nucleic acid
 - 66µl DEPC H₂O
 - 10µl 10X DNase Buffer
 - 10µl DNase (Promega)
- Incubate for 2hrs @ 37°C
- Phenol/Chloroform extract then Ethanol precipitate.
- Resuspend RNA in a small volume of DEPC H₂O and spec.